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DRAG REDUCTION IN WAVE-SWEPT MACROALGAE: **A**LTERNATIVE STRATEGIES AND NEW PREDICTIONS¹

PATRICK T. MARTONE^{2,5}, LAURIE KOST³, AND MICHAEL BOLLER⁴

²Department of Botany and Biodiversity Research Centre, University of British Columbia, Vancouver, British Columbia V6T 1Z4 Canada; ³Hopkins Marine Station of Stanford University, Pacific Grove, California 93950 USA; ⁴St. John Fisher College, Rochester, New York 14618 USA

- *Premise of the study:* Intertidal macroalgae must resist extreme hydrodynamic forces imposed by crashing waves. How does frond flexibility mitigate drag, and how does flexibility affect predictions of drag and dislodgement in the field?
- *Methods:* We characterized flexible reconfiguration of six seaweed species in a recirculating water flume, documenting both shape change and area reduction as fronds reorient. We then used a high-speed gravity-accelerated water flume to test our ability to predict drag under waves based on extrapolations of drag recorded at slower speeds. We compared dislodgement forces to drag forces predicted from slow- and high-speed data to generate new predictions of survivorship and maximum sustainable frond size along wave-swept shores.
- *Key results:* Bladed algae were generally "shape changers", limiting drag by reducing drag coefficients, whereas the branched alga *Calliarthron* was an "area reducer", limiting drag by reducing projected area in flow. Drag predictions often underestimated actual drag measurements at high speeds, suggesting that slow-speed data may not reflect the performance of flexible seaweeds under breaking waves. Several seaweeds were predicted to dislodge at similar combinations of velocity and frond size, suggesting common scaling factors of dislodgement strength and drag.
- *Conclusions:* Changing shape and reducing projected area in flow are two distinct strategies employed by flexible seaweeds to resist drag. Flexible reconfiguration contributes to the uncertainty of drag extrapolation, and researchers should use caution when predicting drag and dislodgement of seaweeds in the field.

Key words: algae; biomechanics; dislodgement; drag coefficient (C_d); evolution; intertidal; macrophyte; morphology; reconfiguration; seaweed.

Waves crashing on shore impose extraordinary hydrodynamic forces on intertidal organisms (Denny, 1988, 1994; Denny et al., 2003). Wave-induced forces affect the survivorship, distribution, and interactions of intertidal animals and algae, thereby influencing population dynamics (Paine, 1979; Blanchette, 1996), zonation patterns (Lewis, 1968; Harley and Helmuth, 2003; Harley and Paine, 2009), and community structure (Connell, 1972; Denny and Wethey, 2001), and making the intertidal zone an excellent test bed for evolutionary and ecological studies (e.g., Dayton, 1975; Sousa, 1979; Paine and Levin, 1981; Harley, 2003; Stachowicz et al., 2008). For example, seaweeds clinging to intertidal rocks must resist wave-induced hydrodynamic forces to survive, just as some terrestrial plants must resist strong winds (Vogel, 1989; Ennos, 1997, 1999; Niklas and Speck, 2001; Butler et al., 2012). Seaweeds that are broken, dislodged, and cast ashore after big storms are testaments to the selective pressures applied by breaking waves. Because seaweeds are foundational species that comprise both food and habitat for animals along the shore, loss of seaweed populations due to wave-induced forces can have cascading effects on intertidal

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⁵Author for correspondence: (e-mail: pmartone@mail.ubc.ca)

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marine communities. Thus, understanding biomechanical adaptations of seaweeds to reduce or resist hydrodynamic forces will help us predict patterns of dislodgement and shifts in nearshore ecology and potentially provide insight into aerodynamic influences on terrestrial plants (Denny, 1994; Ennos, 1999).

Biomechanists have applied engineering principles to quantify hydrodynamic forces experienced by intertidal seaweeds to resolve differences across morphologies and to predict patterns of dislodgement along the shore (Koehl, 1986; Carrington, 1990; Gaylord et al., 1994; Gaylord and Denny, 1997; Hurd, 2000; Denny and Gaylord, 2002; Boller and Carrington, 2006b, 2007; Mach et al., 2011). Among fluid forces, drag has received the most attention, although forces resulting from wave impingement and seaweed inertia have also been demonstrated (Gaylord, 2000; Gaylord et al., 2008). According to previous studies, seaweeds of widely varying morphology often perform similarly in flow because they are flexible. Unlike rigid engineering shapes, such as cones and cylinders, flexible seaweeds "reconfigure" in flow: blades curl up and branches collapse and fold together as water velocity increases, thereby reducing the surface area of fronds projected into the flow and changing shape to limit drag (Carrington, 1990; Denny and Gaylord, 2002; Harder et al., 2004; Boller and Carrington, 2006b).

Boller and Carrington (2006b, 2007) were the first to distinguish between shape change and area reduction in flow, the two drag-limiting processes associated with flexible reconfiguration. Their data demonstrate that some seaweed morphologies are better at reducing projected area in flow, while others are better at reducing drag coefficient (C_d , a parameter that varies with shape). This distinction suggests two morphological strategies exist for limiting imposed drag force: the capacity of flexible

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thalli to change shape or to reduce size. If selection can act on either shape or size changes, then two flexible seaweeds of widely divergent sizes or shapes may ultimately experience similar drag under crashing waves. For example, bladed and branched seaweeds often coexist along the shore, suggesting that either form is sufficient to resist drag. Furthermore, seaweeds that are morphologically similar but phylogenetically distant, such as bladed algae in the Rhodophyte genus Porphyra, the Chlorophyte genus Ulva, and the Phaeophycean genus Petalonia, likely reflect parallel evolution of morphology perhaps canalized in response to selective pressures related to waveinduced drag. Exploring the dynamics of size and shape change of intertidal seaweeds in flow may reveal potential strategies underlying morphological divergence and canalization and may help us understand the evolutionary pressures that gave rise to the diversity of macroalgae along wave-swept rocky shores.

Flexible reconfiguration complicates predictions of drag and dislodgement in the field, as seaweeds flop, twist, and reconfigure in flow. Drag measurements recorded in the laboratory in recirculating flumes (<5 m·s⁻¹) must be extrapolated to environmentally relevant water velocities (up to 25 m·s⁻¹) (Denny and Gaylord, 2002; Denny et al., 2003; Helmuth and Denny, 2003). This extrapolation process is fraught with potential error largely because of uncertainty in reconfiguration (C_d) at high speeds (Vogel, 1994; Bell, 1999). Past studies have reported wide discrepancies between drag predictions made in the laboratory and observations of dislodgement in the field (Gaylord et al., 1994), although some of this error was likely due to a lack of consideration for fatigue of algal tissues under breaking waves (Mach et al., 2007, 2011; Mach, 2009). One notable exception was a recent study by Martone and Denny (2008) that accurately predicted the maximum size of intertidal seaweeds by characterizing drag and dislodgement in a high-speed gravityaccelerated water flume, which generated flow speeds up to 10 m·s⁻¹. The accuracy of their predictions suggests that highspeed measurements may increase our predictive power by reducing the need for extrapolation. But what speeds are sufficient to make accurate predictions?

In this study, we investigate drag and flexible reconfiguration of six morphologically distinct seaweeds in flow. We characterize size and shape change as water velocity increases to explore morphological strategies to mitigate selective pressures applied by wave-induced drag forces. We measure drag and reconfiguration of seaweeds in a recirculating flume up to 4 m·s⁻¹, and then examine the error involved in extrapolation by testing our drag predictions at 6.8 and 9.5 m·s⁻¹ in the high-speed water flume described in Martone and Denny (2008). Finally, we generate updated models of drag and dislodgement for each seaweed species based on all drag measurements up to 9.5 m·s⁻¹, and we test these new models by comparing predicted and observed limits to maximum frond sizes in the field.

MATERIALS AND METHODS

Specimen collection—Seaweeds representing a range of branched and bladed morphologies were collected from the intertidal zone at Hopkins Marine Station, Pacific Grove, California. Fronds (*N* = 10) were collected from each of the following species: *Calliarthron cheilosporioides* Manza, *Chondracanthus exasperatus* (Harvey & J.W.Bailey) J.R.Hughey, *Mazzaella flaccida* (Setchell & N.L.Gardner) Fredericq, *Mastocarpus papillatus* (C.Agardh) Kützing, *Prionitis lanceolata* (Harvey) Harvey, and *Codium fragile* (Suringar) Hariot. Seaweeds were examined and epiphytes were removed. Seaweeds were maintained in running seawater for less than 2 d before experimentation.

Quantifying reconfiguration—Seaweeds were affixed to a calibrated 2-axis force transducer (US-6002, Bokam Engineering, Santa Ana, California, USA) using cyanoacrylate glue. The force transducer was then secured into a recirculating water flume with the seaweed standing erect, perpendicular to the direction of flow. Drag was measured on each seaweed at the following 15 velocities: 0, 0.1, 0.2, 0.3, 0.5, 0.8, 1.2, 1.5, 1.9, 2.3, 2.7, 3.1, 3.4, and 4.0 m·s⁻¹. Free stream velocities were calibrated using an acoustic Doppler velocimeter (ADV Vectrino, Nortek, Rud, Norway). At each velocity, reconfiguring seaweeds were photographed (EOS 30D, Canon) from downstream (see Fig. 1), and seaweed projected areas (A_{proj} were calculated from the average area measured from three photographs using the program ImageJ (National Institutes of Health, Washington, D.C., USA). After each trial, seaweeds were spread out flat and photographed from above to measure maximum area (A_{max} , equivalent to half the wetted surface area).

True drag coefficient ($C_{d, true}$) was calculated at each velocity according to the following equation:

$$C_{\rm d, \, true} = \frac{2 F_{\rm drag}}{\rho U^2 A_{\rm proj}},\tag{1}$$

where F_{drag} is drag force (N), ρ is density of water (1000 kg·m⁻³), *U* is water velocity (m·s⁻¹), and A_{proj} is seaweed projected area (m²) measured from downstream as described above. Differences in true drag coefficients at 4 m·s⁻¹ were detected using a nested two-way ANOVA with factors morphology (2 levels: bladed and branched) and species (6 levels, nested within morphology). Species differences were compared using Tukey's honestly significant difference (HSD) posthoc.

Normalized area $(A_{\%})$ was calculated at each velocity according to the following equation:

$$A_{\%} = \frac{A_{\text{proj}}}{A_{\text{max}}} \times 100.$$
 (2)

Differences in normalized areas at $4 \text{ m} \cdot \text{s}^{-1}$ were detected using a nested twoway ANOVA with factors morphology and species, as defined above. Species differences were compared using Tukey's honestly significant difference (HSD) posthoc.

True drag coefficient ($C_{d, true}$) and normalized area ($A_{\%}$) were plotted against water velocity to provide dimensionless indices of changes in seaweed shape and size with increasing velocity. The product of true drag coefficient and normalized area ($C_{d, true}A_{\%}$) was plotted against water velocity to examine total reconfiguration, the combined effect of both size change and shape change with increasing velocity. Differences in total reconfiguration ($C_{d, true}A_{\%}$) at 4 m·s⁻¹ were detected using a nested two-way ANOVA with factors morphology and species, as defined above. Species differences were compared using Tukey's HSD post hoc test.

Predicting drag at higher velocities—To facilitate extrapolations of drag, we combined the effect of water velocity and frond size into frond Reynolds number (Re_f):

$$\operatorname{Re}_{\mathrm{f}} = \frac{UL}{v} = \frac{U\sqrt{A_{\max}}}{v},\tag{3}$$

where v is the kinematic viscosity of water $(1 \times 10^{-6} \text{ m}^2 \cdot \text{s}^{-1})$ and *L* is the characteristic length (m). This Re_f method was first applied successfully by Martone and Denny (2008). New drag coefficients ($C_{d, \text{planform}}$), based on maximum frond area, were calculated for each seaweed at frond Reynolds numbers tested in the recirculating water flume.

$$C_{\rm d, planform} = \frac{2F_{\rm drag}}{\rho U^2 A_{\rm max}} = \frac{2F_{\rm drag}}{\rho (v \, {\rm Re}_{\rm f})^2}.$$
(4)

For each species, $C_{d, planform}$ was plotted against Re_f, and power curves were fitted to log-transformed data using the program Matlab (v7.0.1, The Mathworks, Natick, Massachusetts, USA), and 95% confidence intervals (CI) around the fitted curves were calculated from 1000 bootstrapped data sets each composed of 10 data points haphazardly sampled from the original data and each data point representing a unique plant and velocity to ensure independence (Efron and Tibshirani, 1993). Estimates of $C_{d, planform}$ (mean ± 95% CI) were then used to estimate drag force up to log Re_f = 6.5, according to the following equation:

$$F_{\rm drag} = \frac{1}{2} \rho U^2 A_{\rm max} C_{\rm d, planform}$$

= $\frac{1}{2} \rho (\nu \operatorname{Re}_{\rm f})^2 C_{\rm d, planform}$ (5)



Fig. 1. Seaweeds reconfiguring in flow, viewed from downstream.

To test these drag predictions, we collected additional fronds: *Calliarthron cheilosporioides* (N = 10), *Chondracanthus exasperatus* (N = 10), *Mazzaella flaccida* (N = 10), *Mastocarpus papillatus* (N = 10), *Prionitis lanceolata* (N = 12), and *Codium fragile* (N = 11). Maximum planform areas (A_{max}) of fronds were measured and recorded as described above. Seaweeds were affixed to a custom force transducer using cyanoacrylate glue and then secured into a gravity-accelerated water flume (illustrated in fig. 3 in Martone and Denny, 2008). Drag was measured on each frond in bursts of water moving 6.8 and 9.5 m·s⁻¹, simulating waves crashing onshore. Drag measurements were plotted against frond Reynolds numbers (Re_{r} , calculated from Eq. 3) and visually compared to predictions based on data generated in the recirculating flume.

New predictions of size and survival—For each species, drag measurements collected in the recirculating and gravity-accelerated flumes were combined into a single data set, and new curves were fitted to log-transformed $C_{d, planform}$ vs. Re_r data. Data collected in the gravity flume were weighted to equalize the number of datapoints analyzed from each flume and to ensure that predictive curves were mostly based on high-speed data. Estimates of $C_{d, planform}$ were used to estimate drag force up to log Re_r = 6.5, according to Eq. 5.

Dislodgement forces were recorded for each seaweed species in the field at Hopkins Marine Station. Fronds (N = 20) from *Prionitis, Chondracanthus, Codium,* and *Mazzaella* were pulled from the rock by attaching a clip near the base of the fronds and using a 5000-g spring scale to record dislodgement force parallel to the substratum. Dislodgement forces of *Calliarthron* were reported by Martone (2006), and dislodgement forces of *Mastocarpus* were reported by Kitzes and Denny (2005). Maximum dislodgement forces were noted, and maximum percentiles were calculated assuming normal distributions and using mean and standard deviation values for each species (Sokal and Rohlf, 2001).

Fronds were predicted to break when drag force and dislodgement force were equal, and maximum dislodgement force represented the maximum drag force that fronds could resist. For each species, drag force was set equal to maximum dislodgement force to calculate the critical frond Reynolds number ($Re_{f, crit}$), the combination of frond size and water velocity that would dislodge all fronds. This was done by iteratively solving Eq. 5 for values of Re_{f} until dislodgement force was reached. $Re_{f, crit}$ was then used to calculate water velocities that would dislodge fronds of a given size or, conversely, the maximum size to which fronds could grow without being dislodged at a given velocity (see Eq. 6). Field observations were used to test these predictions.

$$A_{\rm crit} = \left(\frac{\nu {\rm Re}_{\rm f, \, crit}}{U_{\rm crit}}\right)^2 \tag{6}$$

RESULTS

Quantifying reconfiguration—As water velocity increased in the recirculating flume, fronds reconfigured (Fig. 1), and true drag coefficient decreased for all seaweed species (Fig. 2A). Branched algae *Calliarthron* and *Mastocarpus* had the highest $C_{d, true}$ at all test velocities (Fig. 2A). At 4 m·s⁻¹, $C_{d, true}$ values for bladed algae were significantly lower than $C_{d, true}$ for branched algae ($F_{1, 45} = 150.30$, P < 0.001, Table 1). On average, *Calliarthron* had the highest $C_{d, true}$ at 4 m·s⁻¹ (0.54 ± 0.04), whereas *Mazzaella* had the lowest $C_{d, true}$ (0.08 ± 0.02) ($F_{4, 45} = 15.16$, P < 0.001, Tukey's HSD results in Table 1).

As water velocity increased, normalized area also decreased for all seaweed species (Figs. 1, 2B). Normalized area of *Calliarthron* fronds decreased more than that of other seaweeds. At 4 m·s⁻¹, normalized area of *Calliarthron* decreased to $6.2 \pm 0.5\%$ of maximum frond area, more area reduction than all other seaweeds investigated ($F_{4,45} = 24.29$, P < 0.001, Tukey's HSD results in Table 1). At 4 m·s⁻¹, normalized areas of other seaweeds decreased to 14.9–22.9%, on average, and did not differ significantly by morphology ($F_{1,45} = 1.30$, P = 0.26) or by species (Tukey's HSD results in Table 1).

The combination of true drag coefficient and normalized area also decreased with increasing water velocity (Fig. 2c). In general, bladed algae reconfigured significantly more than branched algae ($F_{1,45} = 68.53$, P < 0.001). Mazzaella reconfigured significantly more than all other seaweeds ($F_{4,45} = 10.87$, P < 0.001, Tukey's HSD results in Table 1). Considering both shape and area change, reconfiguration of *Calliarthron* fronds was comparable to other seaweeds (Fig. 2C, Tukey's HSD results in Table 1).

Predicting drag at higher velocities—Experiments in the recirculating water flume generated drag data in the range of log Re_f from approximately 3.5–5.5 (Fig. 3). For all species, recalculated drag coefficients ($C_{d, planform}$) decreased as Re_f increased (Fig. 3). Curves fitted to $C_{d, planform}$ vs. Re_f data generally had little margin of error (Fig. 3); however, extrapolations of drag force beyond the data often had wide margins of error and were not always accurate (Fig. 4). For *Codium, Prionitis, Mastocarpus*, and *Chondracanthus*, data collected in the recirculating water flume were reasonably successful in predicting drag at higher Re_f (Fig. 4B–E). However, for *Calliarthron* and *Mazzaella*, data collected in the recirculating flume often under-predicted drag experienced by fronds at higher Re_f (Fig. 4A, F). Predictions for *Mazzaella* were particularly poor; drag forces on *Mazzaella* fronds were many times higher than those predicted (Fig. 4F).

Predicting dislodgement—With increasing Re_f, drag on *Codium* and *Mastocarpus* was predicted to increase similarly and at a faster rate than drag on other seaweeds (Fig. 5A). Drag on *Chondracanthus* and *Prionitis* was also predicted to increase similarly (Fig. 5A). Among the seaweeds studied, drag on *Mazzaella* blades was predicted to increase most slowly with increasing Re_f (Fig. 5A).

Maximum dislodgement forces of the six seaweed species are listed in Table 2. Maximum forces ranked above the 89th percentile of dislodgement forces for each species (Table 2), suggesting that these forces would be sufficient to dislodge at least 89% of the individuals of each species. Maximum dislodgement forces were used in conjunction with drag prediction curves to calculate Ref, crit values (Table 2, Fig. 5A). Ref, crit values were similar (approximately 2×10^6) for four wave-exposed species: Prionitis, Calliarthron, Chondracanthus, and Maz*zaella* (Table 2, Fig. 5A). Re_{f,crit} values of *Codium* (0.989×10^6) and *Mastocarpus* (0.479×10^6) were lower than those of the other four species. Ref, crit values generated isoclines for each species depicting decreasing maximum frond size with increasing velocity (Fig. 5B). Larger fronds were predicted to break at slower velocities; smaller fronds were predicted to break at faster velocities (Fig. 5B). Isoclines for Prionitis, Calliarthron, Chondracanthus, and Mazzaella were similar, and for any water velocity these four species were predicted to support larger fronds than Codium or Mastocarpus (Fig. 5B). The maximum observed size of Mazzaella and Chondracanthus were similar and were predicted to resist similar water velocities (Fig. 5B). The maximum observed size of Mastocarpus and Calliarthron were similar (approximately 40 cm²), but at this size Calliar*thron* was predicted to resist nearly four times the water velocity before breaking (Fig. 5B).

DISCUSSION

Hydrodynamic selective pressures—During reconfiguration, shape change and area reduction occur concurrently, yet they are somewhat independent of one another: seaweeds that quickly reduce projected areas may maintain relatively high drag coefficients, and seaweeds that are exceptionally good at reducing drag coefficients may maintain large projected areas. In other words, seaweeds specializing in area reduction or shape change may experience similar drag despite morphological differences. Thus, as seaweeds have evolved in hydrodynamically stressful habitats like the wave-swept intertidal zone, selection may have acted on the ability of macroalgal thalli to either change shape or to reduce area in flow.

Our data illustrate these two strategies for drag limitation. For example, drag coefficients of the articulated coralline *Calliarthron* are consistently higher than drag coefficients of all other algae in this study, suggesting that its calcified, segmented structure is particularly drag prone. However, *Calliarthron* fronds compensate for high drag coefficients by being exceptionally good at reducing projected area as flow rates increase. When both shape change and area reduction are taken into account, total reconfiguration of *Calliarthron* fronds is intermediate and not unlike other seaweeds (Fig. 2C). Thus, *Calliarthron* is an "area reducer," limiting drag primarily by reducing frond area projected into flow, not by changing shape.

Bladed algae, on the other hand, are "shape changers." Both *Chondracanthus* and *Mazzaella* have significantly lower drag coefficients than branched algae at the highest experimental velocities (Fig. 2A). However, their capacity to reduce projected area in flow was comparable to other branched algae. When shape change and area reduction are taken into account, *Chondracanthus* and *Mazzaella* reconfigured more than most seaweeds, and *Mazzaella* significantly so. As "shape changers," *Mazzaella* and *Chondracanthus* limit drag primarily by reducing drag coefficient, not necessarily by reducing projected area. The slight advantage of blades to limit drag may help explain how many of the largest seaweeds, such as *Durvillaea* spp., *Saccharina* spp., and *Pleurophycus* sp., are able to support this morphology in hydrodynamically stressful habitats.



Fig. 2. Quantifying shape change and area reduction of seaweed thalli with increasing water velocity. (A) Reduction in true drag coefficient ($C_{d, true}$). (B) Reduction of projected area ($A_{\%}$). (C) Combined reduction of true drag coefficient and projected area ($C_{d, true} \cdot A_{\%}$).

Species	$C_{ m d, true}$	$A_{\it m}$	$C_{ m d,\ true}{\cdot}A_{\%}$	Morphology
Calliarthron	0.54 ± 0.04 a	6.2 ± 0.5 a	0.033 ± 0.002 bc	branched
Codium	$0.24 \pm 0.01 \text{ b}$	22.9 ± 0.9 bc	0.054 ± 0.004 ab	branched
Mastocarpus	0.37 ± 0.02 a	22.8 ± 3.1 bcd	0.085 ± 0.043 a	branched
Prionitis	0.20 ± 0.02 b	$14.9 \pm 1.1 \text{ bd}$	0.029 ± 0.005 c	branched
Chondracanthus	0.13 ± 0.03 c	19.2 ± 1.2 bcd	0.024 ± 0.004 c	blade
Mazzaella	$0.08 \pm 0.02 \text{ c}$	18.8 ± 7.6 bcd	$0.013 \pm 0.003 \text{ d}$	blade

TABLE 1. Drag coefficients and projected areas of each species at 4 m·s⁻¹.

Notes: $C_{d,true}$ = true drag coefficient, $A_{\%}$ = normalized projected area, letters within each column indicate differences among species based on post-hoc Tukey's tests (P < 0.05).

Certainly, these two strategies for drag limitation are not exclusive of one another; some algae likely employ both strategies equally. For example, across all test velocities, the branched alga *Prionitis* exhibits intermediate drag coefficients, intermediate projected areas and, indeed, intermediate levels of total reconfiguration. Thus, *Prionitis* is not clearly an "area reducer" or a "shape changer". Nevertheless, distinguishing between these two adaptive strategies for reducing drag will help us understand the selective pressures influencing the morphological evolution of wave-swept seaweeds. Unfortunately, both shape change and area reduction are dynamic processes that are difficult to predict once seaweeds are collected and removed from natural flow conditions. Selection must act on "the ability to change shape" or "the ability to reduce projected area," and these two abilities are not obviously linked to static algal morphologies. Indeed, under natural flow conditions, seaweeds of differing morphology all start to look similar (see Fig. 1). As past studies have explored the interaction between material properties and drag (Koehl, 1986; Denny et al., 1997; Gaylord and Denny, 1997; Koehl, 2000; Boller and Carrington, 2007; Demes et al., 2011), future studies may find value in material properties, such as Young's



Fig. 3. Drag coefficient ($C_{d, planform}$) vs. frond Reynolds number (Re_f) for each experimental species. Open circles are data collected in the recirculating flume, and gray regions represent ±95% confidence intervals around predictions based on flume data alone. Black triangles are data collected in the high-speed gravity flume; solid lines are curves fitted to both recirculating and gravity flume data, as described in the methods.



Fig. 4. Drag force (F_{drag}) vs. frond Reynolds number (Re_f) for each experimental species. Open circles are data collected in the recirculating flume, and gray regions represent ±95% confidence intervals around predictions based on flume data alone. Black triangles are data collected in the high-speed gravity flume; solid lines are curves fitted to both recirculating and gravity flume data, as described in the methods.

modulus or flexural stiffness, to predict the ability of seaweeds to change shape or projected area in flow.

It should be noted that $C_{d, true}$ values documented at slowspeed (Fig. 2A, $U = 0.1 \text{ m} \cdot \text{s}^{-1}$) were notably high for some seaweeds, such as *Calliarthron* and *Mastocarpus*—up to 10 times greater than those reported for various engineering shapes (Vogel, 1994), which rarely exceed 1 at a comparable Reynolds number (>10³). The cause of these high drag coefficients is unknown and deserves further study. It may be a consequence of shear drag on irregularly shaped fronds at slow speeds or the result of fronds flapping to form unexpectedly large wakes in flow (Hoerner, 1965; Koehl and Alberte, 1988).

Predicting drag at high velocities—Past studies have had difficulty linking drag measured in slow-speed laboratory conditions to observed patterns of survivorship in the field. Blame is often focused on the notoriously inaccurate process of drag extrapolation. Indeed, in the current study, data collected on seaweeds in the recirculating flume did not always accurately predict drag at higher speeds. Predicted and observed drag forces at higher speeds were similar for only some of the seaweeds studied here (*Prionitis, Codium, Mastocarpus*, and

Chondracanthus), suggesting that measurements at slower speeds might be sufficient to predict performance of these seaweeds in the field. However, for the other seaweeds studied here (*Mazzaella* and *Calliarthron*), drag predictions underestimated actual drag measurements at high speeds, suggesting that slow-speed data collected on these seaweeds in the laboratory are misleading and are not representative of their performance in the field. That 33% of drag extrapolations were inaccurate supports and highlights the uncertainty noted in previous studies and underscores the need for researchers to think twice before drawing conclusions based on extrapolations.

The discrepancy between predicted and observed drag forces at high speeds suggests that, at least for some seaweeds, slow-speed reconfiguration in recirculating flumes may differ from high-speed reconfiguration under breaking waves. In recirculating flow, seaweeds have more time to reconfigure, bending and collapsing their branches to a greater extent as flow steadily increases; whereas, under breaking waves (and in the high-speed flume here), seaweeds are suddenly struck by incoming waves, making them susceptible to wave-impingement forces (Gaylord et al., 2008) and allowing only minimal time to reconfigure and limit drag. This difference in time-dependent



Fig. 5. Drag, maximum size, and critical water velocity predictions for seaweed thalli in the field. (A) Drag force (F_{drag}) predictions vs. frond Reynolds number (Re_f). Shapes represent maximum dislodgement forces, used to calculate critical Re_f. Dotted lines represent hypothetical dislodgement forces greater than the strongest alga observed. (B) Critical Re_f isoclines for each species. Shapes represent maximum frond sizes (A_{crit}) observed in the field, used to predict the maximum water velocity (U_{crit}) experienced by each species along the shore. Dotted lines represent hypothetical planform areas greater than the largest alga observed.

reconfiguration would explain why drag forces at higher speeds were greater than those predicted by slow-speed data (e.g., see Fig. 4F). Unfortunately, turbulent and aerated flow conditions make it impossible to photograph seaweed reconfiguration under breaking waves, preventing a complete analysis of shape change and area change at high speeds. Thus, we do not know if extrapolation error is due to

TABLE 2. Maximum observed frond sizes, dislodgement forces, and critical frond Reynolds numbers (Re_{f.crit}) for each species.

		Dislodgement force (N)			
Species	Max observed planform area (cm ²)	Mean ± SD	Max	Max percentile	$\begin{array}{c} \text{Re}_{\text{f,crit}} \\ (\times 10^6) \end{array}$
Calliarthron	41.0	18.5 ± 5.1	24.5	89.4	1.762
Codium	137.0	17.1 ± 4.6	22.5	89.4	0.989
Mastocarpus	46.0	3.4 ± 1.4	6.6	98.7	0.479
Prionitis	161.5	19.1 ± 7.3	37.8	99.4	2.006
Chondracanthus	222.1	20.9 ± 6.3	34.1	98.4	1.907
Mazzaella	210.0	12.1 ± 4.1	19.6	96.7	1.972

time-dependent reconfiguration or some other hydrodynamic phenomenon.

Predicting dislodgement—New drag predictions based on both slow- and high-speed data show that seaweed species group together in surprising ways. As velocity and size increase (i.e., Re_f increases), drag is predicted to increase similarly and most rapidly for thalli of *Codium* and *Mastocarpus*, two seaweeds of widely divergent branching structure and thallus construction (Fig. 5A). Perhaps more perplexing is that drag is predicted to increase similarly on branched thalli of *Prionitis* and bladed thalli of *Chondracanthus*. These data suggest that comparisons of drag experienced by bladed and branched algae may not be generalizable.

In the field, drag forces are resisted by dislodgement forces, which differ among seaweed species. Although maximum dislodgement forces varied widely among experimental seaweeds, many of these values point to strikingly similar Ref. crit values that would lead to breakage. Wave-exposed thalli of *Prionitis*, Calliarthron, Chondracanthus, and Mazzaella are all predicted to fail when $\operatorname{Re}_{f, \operatorname{crit}}$ approaches 2×10^6 . This suggests that attachment strength and drag force may scale similarly in these wave-exposed taxa. For example, Mazzaella experiences relatively low drag but also has a low maximum attachment strength; Prionitis experiences more drag, but is stronger. Drag-related scaling patterns such as these are likely to exist in nature, since drag resistance depends critically upon the mechanical properties (e.g., strength) of supporting seaweed tissues (see Martone, 2007). The identification of common Ref. crit values among morphologically distinct wave-swept taxa is interesting and deserves further study. Is there something special about this combination of velocity and frond size?

Because Ref, crit values were comparable among Prionitis, Calliarthron, Chondracanthus, and Mazzaella, fronds of a given size were all predicted to resist similar water velocities. As fronds grow, slower water velocities are required to dislodge them. Therefore, the isoclines depicted in Fig. 5B can be used in combination with maximum observed frond sizes to explore the hydrodynamic environment where experimental seaweeds were collected. For example, the largest thalli collected in this study were blades of Mazzaella and Chondracanthus. Ref. crit isoclines suggest that these large fronds could not have experienced water velocities greater than 15 m·s⁻¹, setting an upper bound to the likely wave exposure at that intertidal site. On the other hand, the maximum size of Calliarthron fronds was much smaller, suggesting that intertidal Calliarthron fronds may have experienced up to 28 m·s⁻¹. These estimates are in general agreement with past measurements of intertidal water velocities (Denny et al., 1989, 2003; Denny, 1994; Gaylord et al., 1994; Denny and Gaylord, 2002; Martone and Denny, 2008; Mach et al., 2011). However, fronds that exceed the maximum sizes observed in this study may be found growing in the subtidal zone or in areas of reduced wave exposure.

In general, smaller fronds are not as limited by water velocities, and so it is possible that young or small mature fronds may be found growing at more wave-battered locations. Our model lacks consideration of the relationship between size and attachment strength; if small plants have lower attachment strength (e.g., Gaylord et al., 1994), then the water velocity predicted to break small fronds in Fig. 5B may be an overestimate. On the other hand, some red algae exhibit little variation in attachment strength across size classes (Carrington, 1990; Shaughnessy et al., 1996; Kitzes and Denny, 2005; Martone, 2006, 2007), so the current model may be reasonable.

The Re_{f, crit} isoclines presented here can be used to make broad comparisons across seaweed species and habitats. For a given size, fronds of Codium and Mastocarpus are predicted to break at lower water velocities than fronds produced by the other four species. This suggests that these two species may persist at wave-exposed locations, but only if they remain relatively small. The maximum observed sizes of Mastocarpus and Calliarthron were similar, but at this size, Mastocarpus could only resist one-quarter the critical water velocity of Calliarthron before being dislodged. Why the discrepancy? Perhaps Mastocarpus fronds living higher in the intertidal zone do not experience the same rapid water velocities experienced by Calliarthron and other wave-exposed algae growing in the low intertidal zone. Another possibility is that Mastocarpus fronds growing in tight clusters resist faster water velocities by achieving a "drafting" benefit from neighboring fronds and thereby experience less drag (Johnson, 2001; Boller and Carrington, 2006a). Similarly, the maximum size of Codium fronds was almost as large as other wave-swept algae, but such large fronds were predicted to break at less than 10 m·s⁻¹ water velocity. Large Codium fronds are commonly found in sheltered marinas where water velocities are slow, but fronds collected for this study were collected from a generally wave-exposed shore. Perhaps these wave-exposed Codium fronds survived by experiencing locally reduced flow conditions in tidepools or in the lee of big rocks. Understanding the hydrodynamic limits of intertidal seaweeds may provide an additional index of wave exposure and thereby improve predictions of local water velocities along waveswept shores (Helmuth and Denny, 2003).

In summary, intertidal seaweeds limit wave-induced drag forces by changing shape (i.e., reducing drag coefficient) and by reducing area projected in flow. These two drag-limiting processes are intertwined but distinct and likely play a role in the morphological evolution of wave-swept seaweeds. Flexible reconfiguration permits certain bladed and branched algae to perform similarly in flow, despite morphological differences, and often complicates our ability to predict drag and dislodgement of seaweeds in the field.

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